



**Providing trust**

## LABORATORY ANALYSIS REPORT

for

**THE HEALTH COMMISSION,  
KENYA CONFERENCE OF CATHOLIC  
BISHOPS  
NAIROBI**

Date of Sampling:  
Date of Report:

13<sup>th</sup> November 2014  
16<sup>th</sup> November 2014

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### **3.0 INTRODUCTION OF THE COMMISSION AND METHODS**

The following document details the methods used:

#### **3.1.1 To evaluate the overall KCCB performance in relation to the delivery of national environmental health services in Kenya.**

3.1.2 To identify and recommend the areas health commission should focus on during the review exercise.

#### **3.2 REFERENCES IDENTIFIED FOR THE REVIEW AND ASSESSMENT**

3.2.1 The following documents were used for reference and assessment purposes:  
Kenya High Level Environmental Documentation

3.2.2 Kenya Health Sector Strategy, April 2009; Guidelines for the assessment of the performance distribution of health interventions in Kenya, including radiography

#### **3.3 CONSTITUTION OF THE ASSESSMENT TEAM**

The assessment was carried out as follows:

#### **3.3.1 Sampling of the implementation methods used in Kenya**

November 2014

3.3.2 Laboratory Analysis of the Kenyan performance data from the 14th quarter

(July to September 2014)

#### **3.3.3 Reporting and presentation of the data and findings**

## **1.0 EXECUTIVE SUMMARY**

The Through the Catholic Health Commission of Kenya, the Catholic Church runs close to 30% of all healthcare facilities in Kenya. The Church has an expansive network which consists of 448 health units (54 hospitals, 83 health centres and 311 Dispensaries) and more than 46 Community Based Health and Orphaned and Vulnerable Children (OVC) Programs. In the Arid and Semi-Arid Areas, the Church has and manages mobile clinics for nomadic communities.

The Catholic Health commission has requested for laboratory analysis of six samples of the tetanus vaccine that was administered by the World Health Organization (WHO) / United Nations Childrens Fund (UNICEF) in the last campaign to determine the presence of human chorionic gonadotropin hormone (HCG).

The scope of the laboratory assessment was to determine the presence of human chorionic gonadotropin hormone (HCG) in all the samples collected.

### **1.1 OBJECTIVES OF THE LABORATORY ASSESSMENT**

The Laboratory assessment had two objectives:

- 1.1.1 To assist the Catholic Health Commission in determining the presence of human chorionic gonadotropin hormone (HCG) in the samples tested.
- 1.1.2 To inform and interpret to the Catholic Health Commission the findings from the laboratory analysis

### **1.2 REFERENCES USED DURING THE LABORATORY ASSESSMENT**

- 1.2.1 Purification of Human Chorionic Gonadotropin Hormone by Anion-Exchange High Performance Liquid Chromatography (HPLC)
- 1.2.2 Journal of Chromatography A, 847 (1999) - Determination of the molecular size distribution of type b tetanus toxoid conjugate vaccines by size exclusion chromatography

### **1.3 EXECUTION OF THE ASSESSMENT**

The assessment was carried out at as follows:

- 1.3.1 Sampling of the samples at St. Michaels Medical Care clinic on the 13<sup>th</sup> of November 2014
- 1.3.2 Laboratory Analysis using High Performance Liquid Chromatography (HPLC) on the 14<sup>th</sup> of November 2015
- 1.3.3 Reporting and interpretation of the data on 15<sup>th</sup> November 2014

## **2.0 SUMMARY OF FINDINGS**

Tetanus is acquired when the spores of the bacterium Clostridium Tetani infect a wound or the umbilical stump. Spores are universally present in the soil. People of all ages can get tetanus but the disease is particularly common and serious in newborn babies ("neonatal tetanus"). It requires treatment in a medical facility, often in a referral hospital. Neonatal tetanus, which is mostly fatal, is particularly common in rural areas where deliveries are at home without adequate sterile procedures. WHO estimated that neonatal tetanus killed about 180 000 babies in 2002.

Tetanus can be prevented through immunization with tetanus-toxoid (TT) -containing vaccines. Neonatal tetanus can be prevented by immunizing women of childbearing age with tetanus toxoid, either during pregnancy or outside of pregnancy. This protects the mother and - through a transfer of tetanus antibodies to the fetus - also her baby.

Human Chorionic Gonadotropin (HCG) hormone is synthesized by the chorionic tissue of the placenta and is found in urine during pregnancy. HCG comprises the glycoprotein hormone family. HCG is dimeric and is composed of two non-covalently bonded glycopeptides sub-units termed  $\alpha$  (alpha HCG) and  $\beta$  (beta HCG).

A High Performance Liquid Chromatography (HPLC) method has been developed for the detection of HCG hormone sample in one chromatographic run using anion exchange chromatography. During the 60 minute linear gradient run, complete separation was accomplished in 40 minutes. The retention time for the HCG peak using this method was about 35 minutes.

Of the six vaccine samples collected and subjected to Laboratory HPLC analysis, the following samples were found to contain the HCG hormone:

- Sample S1 KA retention time 36.283 mins and peak area of HCG was 37%
- Sample S2 KB retention time 35.825 mins and peak area of HCG 26.5%
- Sample S3 KC retention time 36.583 mins and peak area of HCG 23.9%

The laboratory assessment has revealed that three out of the six tetanus vaccine samples analysed contain the human chorionic gonadotropin (HCG) vaccine.

### **3.0 INTRODUCTION**

Human chorionic Gonadotropin (HCG) hormone is synthesized by the chorionic tissue and is found in urine during pregnancy.

HCG together with luteinizing hormone (LH), follicle-stimulating hormone (FSH, follitropin) and thyroid-stimulating hormone (TSH, thyrotropin) comprise the glycoprotein hormone family. All are dimeric and composed of two non-covalently bonded glycopeptide subunits termed  $\alpha$  and  $\beta$ . The total number of amino acid residues in HCG for the  $\alpha$  subunit is 92 and for the  $\beta$  chain is 147. The amino acid sequences of both subunits of HCG have also been determined by Pierce and Morgan et al. Oligosaccharide and sialic acid chains are attached to both subunits. Each subunit is extensively crosslinked by intramolecular disulfide bonds.

A great number of immunological methods for HGC assays are available. These are based on haemagglutination, latex particle agglutination, complement fixation and radioimmuno-reaction. The sensitivity and specificity of these methods make them potentially useful for the measurement of HCG hormone in urine concentration.

High separation of HCG has been obtained by simple methods of column chromatography on diethyl-aminoethyl (DEAE) Sephadex and Sephadex G-100 columns by Bell et al. Another method employing stepwise gradient with increasing NaCl concentration was reported by Yi-Han Chang et al. The isolation of HCG using these methods would be suitable but the procedures are time consuming, the standard column chromatographic procedures taking about two [2] weeks.

A rapid isolation of HCG can be accomplished in one day using anion exchange chromatography.

#### **3.1 Description of the Analytical Method**

The isoelectric point of the HCG molecule is about 4, solutions with higher pH contain HCG molecules in anionic form.

Eluent "A" was phosphate buffer [ph=6], while eluent "B" was 0.01M phosphate buffer [pH=3]. Buffer "B" contained 0.05M sodium sulfate and 0.05M sodium hydrogen sulfate. The pH of the solution was acidic due to the acidic character of the NaHSO<sub>4</sub>. Buffer "A" contained 0.01% v/v of "B" leading to the sulfate and hydrogen ion concentrations being 10,000 times smaller. This means that during a 40 min linear gradient both the hydrogen ion and the sulfate concentrations have been significantly increased, conditions that promote the elution and good resolution of HCG.

The retention time of the HCG peak was about 35mins.

### **3.2 HPLC Apparatus and Columns**

#### **3.2.1 HPLC Equipment**

The analytical HPLC system used in this assessment consisted of a Shimadzu Class VP 10 system. A Dell computer connected to Shimadzu SCL 10A - system controller was used for gradient control of the two shimadzu pumps.

The preparative procedures were accomplished on Shimadzu LC-10AT Liquid Chromatograph connected to a Shimadzu 10AV UV VIS detector with a model 201 fraction collector and equipped with a 7125 sample injection valve with 1ml sample loop.

#### **3.2.2 Column**

Anion exchange separations were performed on an ODS 5 $\mu$ m 250mm x 4.6mm column.

#### **3.2.3 Materials**

Mobile phases contained analytical - grade sodium acetate, sodium bicarbonate, potassium chloride, potassium dihydrogen phosphate, dipotassium hydrogen phosphate, sodium sulfate and sodium hydrogen sulphate.

HPLC grade acetonitrile and deionized water were used for preparing the eluents. Solvents were degassed ultrasonically.

#### **3.2.4 Gradient Programs**

The chromatographic run started isocratically by pumping 100% "A" mobile phase for 20min. A linear gradient from 0% "B" to 100% "B" mobile phase was employed at a 1.0ml/min flow rate for 40 mins. The samples were dissolved in "A" eluent and injected at the beginning of the gradient run.

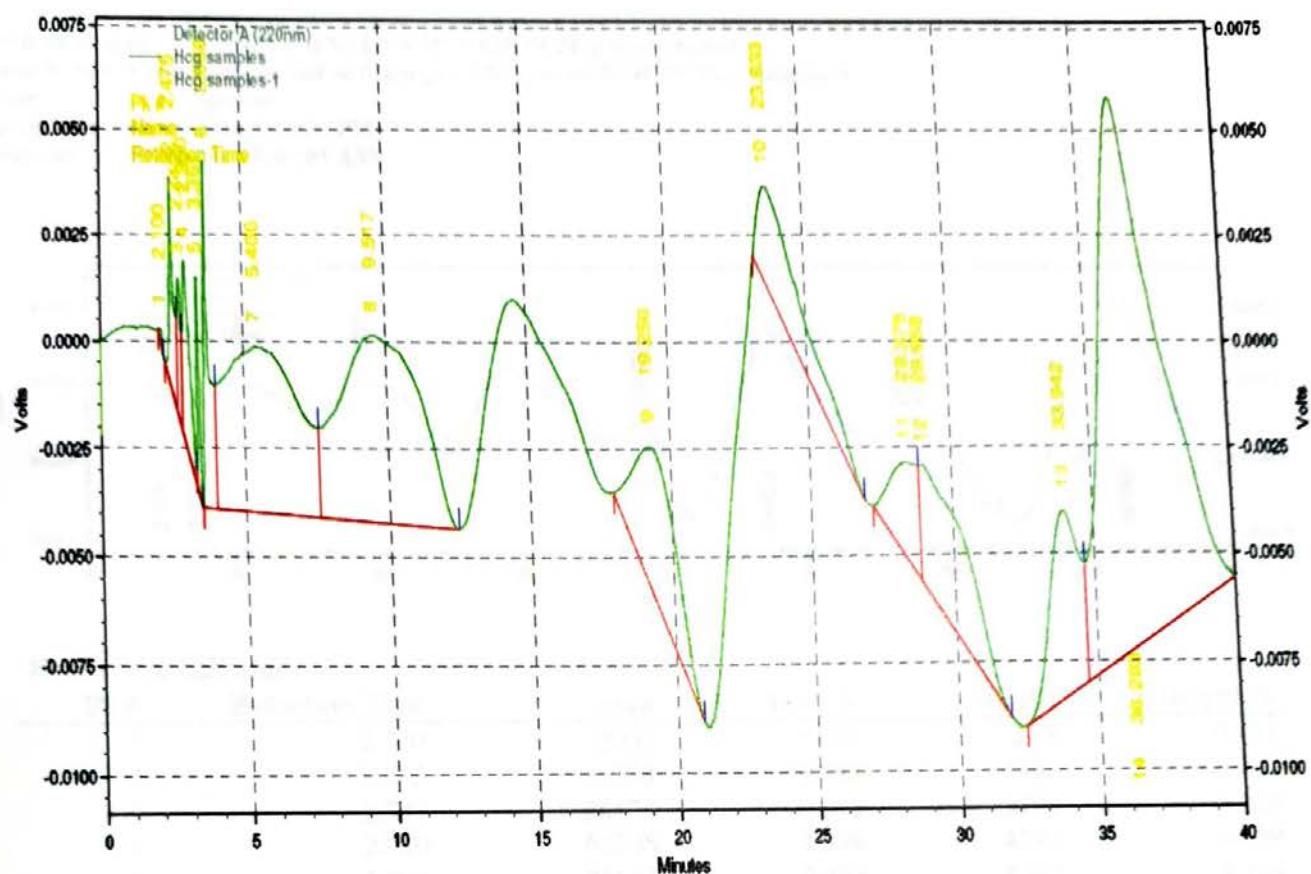
## **4.0 RESULTS**

The samples analysed were:

- Sample 1 - KA
- Sample 2 - KB
- Sample 3 - KC
- Sample 4 - KD
- Sample 5 - KE
- Sample 6 – KF

#### 4.1 Results Sample KA

HPLC chromatogram of Sample KA

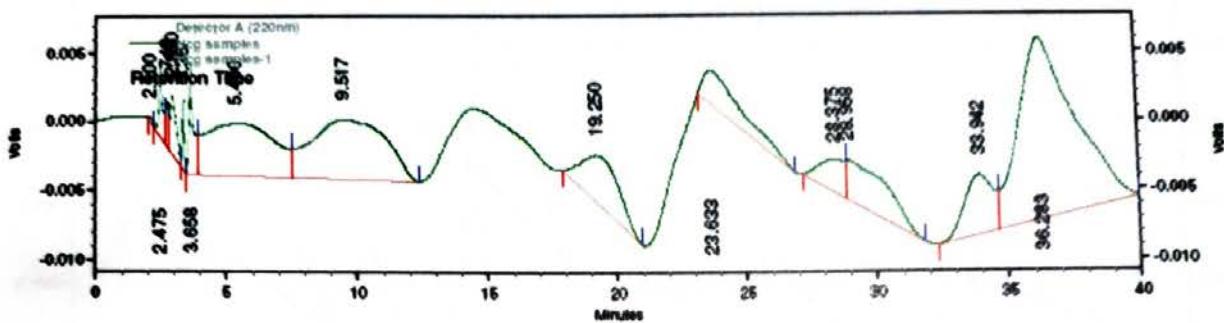


### Peak area Calculation of Sample KA

Shimadzu CLASS-VP V6.12 SP4  
Page 1 of 1

*Area % Report*

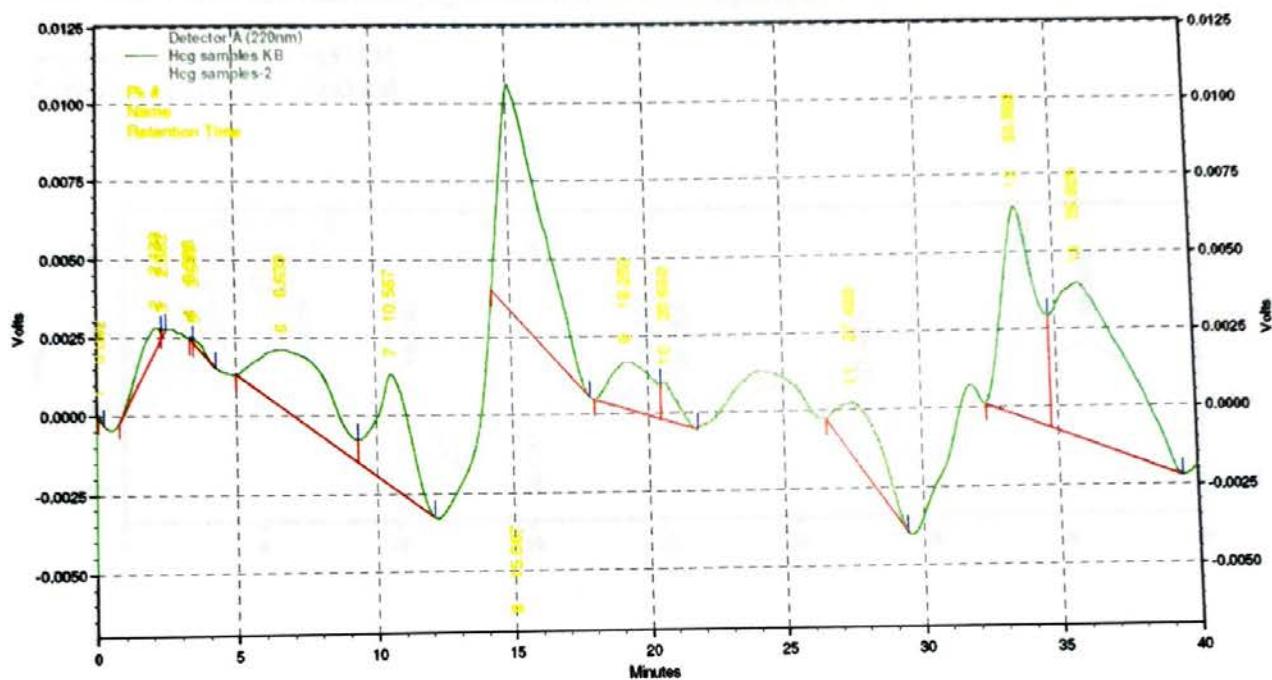
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Pk #	Retention Time	Area	Area %	Height	Height %
1	2.100	2003	0.038	268	0.431
2	2.475	46218	0.888	4982	8.005
3	2.742	28178	0.542	3290	5.286
4	2.950	67598	1.299	4294	6.899
5	3.367	25661	0.493	5042	8.101
6	3.658	98852	1.900	8132	13.066
7	5.400	690049	13.263	3835	6.162
8	9.517	856619	16.464	4316	6.935
9	19.250	351664	6.759	3389	5.445
10	23.633	268812	5.167	2311	3.713
11	28.375	152597	2.933	2257	3.626
12	28.958	344159	6.615	2814	4.521
13	33.942	317414	6.101	4258	6.842
14	36.283	1953152	37.539	13049	20.967
<b>Totals</b>		<b>5202976</b>	<b>100.000</b>	<b>62237</b>	<b>100.000</b>

## 4.2 Results Sample KB

HPLC chromatogram of Sample KB



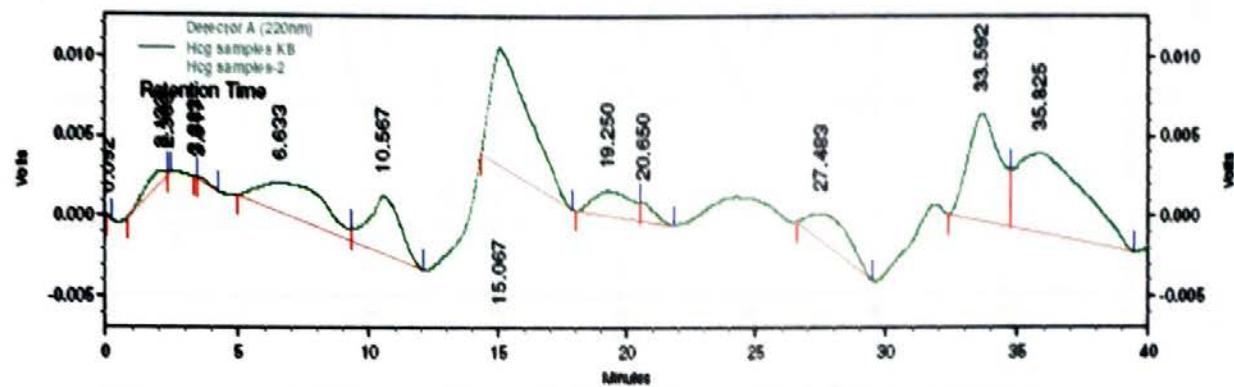
### Peak Area Calculation Sample KB

Shimadzu CLASS-VP V6.12 SP4

Area % Report

Page 1 of 1

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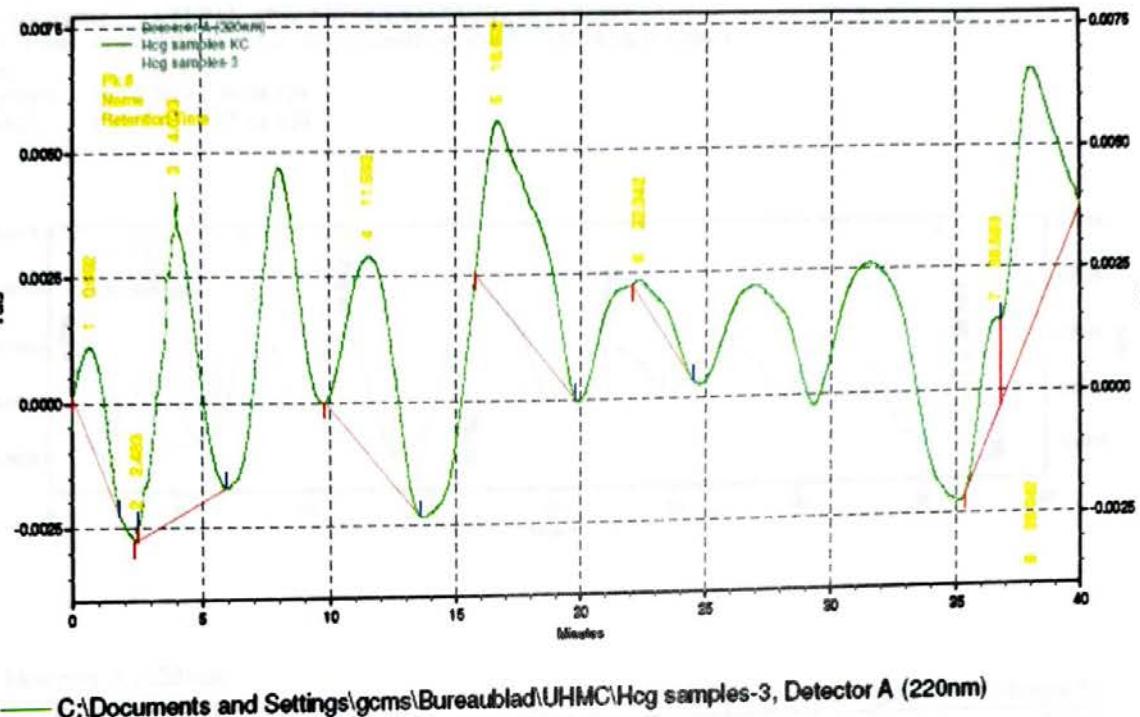


#### Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.092	646	0.019	122	0.403
2	2.133	47648	1.417	720	2.378
3	2.383	1932	0.057	242	0.799
4	3.383	635	0.019	109	0.360
5	3.517	5308	0.158	127	0.420
6	6.633	362484	10.783	1849	6.108
7	10.567	301380	8.965	3615	11.942
8	15.067	768797	22.869	7338	24.240
9	19.250	158761	4.723	1511	4.991
10	20.650	48302	1.437	1216	4.017
11	27.483	204585	6.086	1698	5.609
12	33.592	570014	16.956	6729	22.228
13	35.825	891274	26.512	4996	16.504
Totals		3361766	100.000	30272	100.000

### 4.3 Results Sample KC

HPLC chromatogram of Sample KC



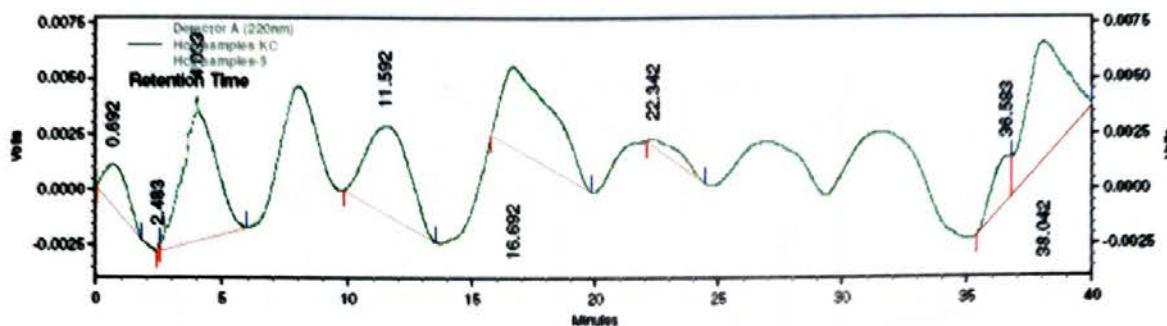
## Peak Area Calculation sample KC

Shimadzu CLASS-VP V6.12 SP4

*Area % Report*

Page 1 of 1

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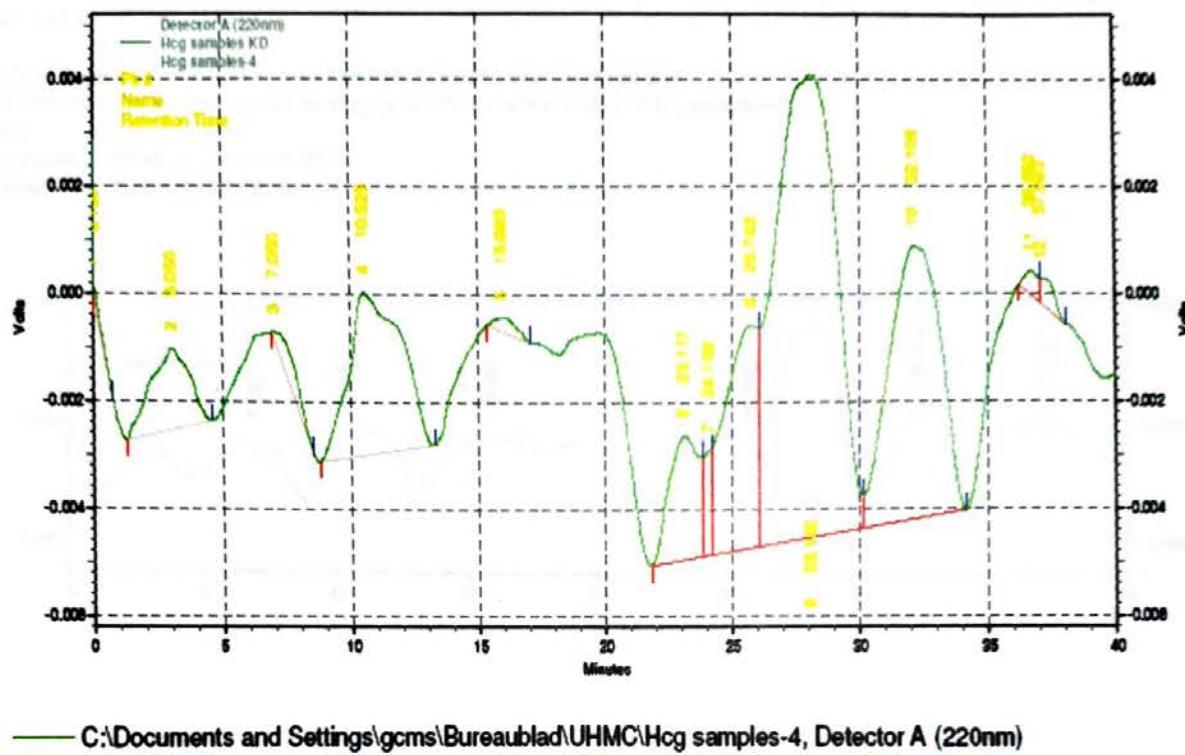


Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.692	125303	5.108	1846	7.740
2	2.483	1405	0.057	315	1.321
3	4.033	555466	22.644	6493	27.223
4	11.592	480145	19.573	3979	16.683
5	16.692	543752	22.166	3605	15.115
6	22.342	50716	2.067	328	1.375
7	36.583	109024	4.444	1993	8.356
8	38.042	587238	23.939	5292	22.188
Totals		2453049	100.000	23851	100.000

#### 4.4 Results Sample KD

HPLC Chromatogram of Sample KD



### Peak Area Calculation Sample KD

Shimadzu CLASS-VP V6.12 SP4

*Area % Report*

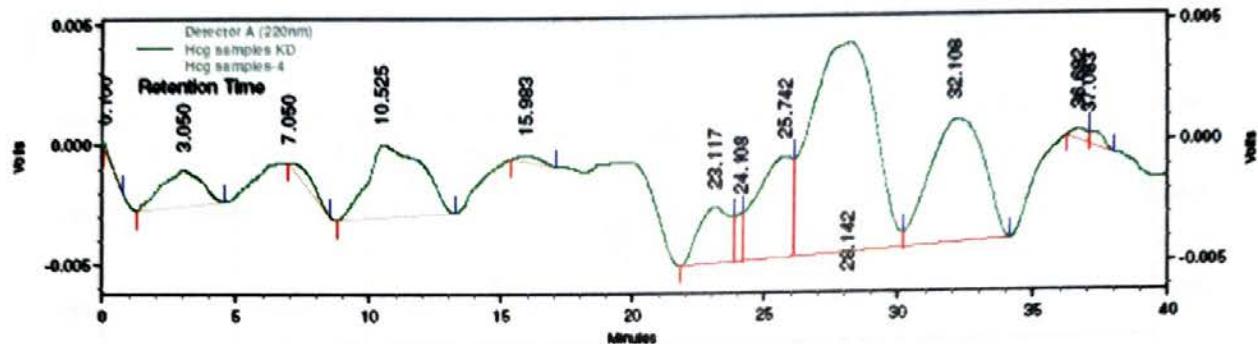
Page 1 of 1

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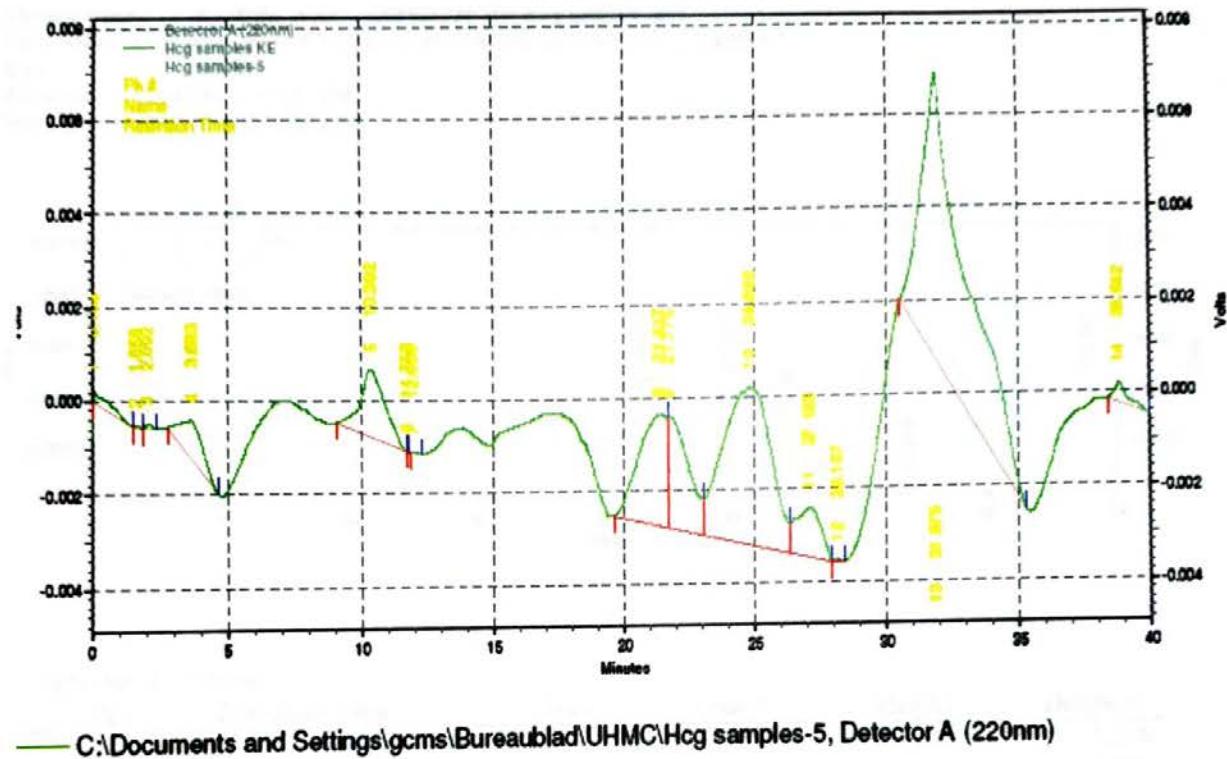


Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.100	4142	0.121	308	1.087
2	3.050	157140	4.586	1494	5.272
3	7.050	31315	0.914	139	0.491
4	10.525	430352	12.560	3042	10.735
5	15.983	16450	0.480	271	0.956
6	23.117	186997	5.458	2321	8.190
7	24.108	39286	1.147	1984	7.001
8	25.742	381357	11.130	4157	14.669
9	28.142	1435337	41.891	8633	30.464
10	32.108	707272	20.642	5072	17.898
11	36.692	16431	0.480	445	1.570
12	37.083	20288	0.592	472	1.666
<b>Totals</b>		<b>3426367</b>	<b>100.000</b>	<b>28338</b>	<b>100.000</b>

#### 4.5 Results Sample KE

HPLC Chromatogram Sample KE



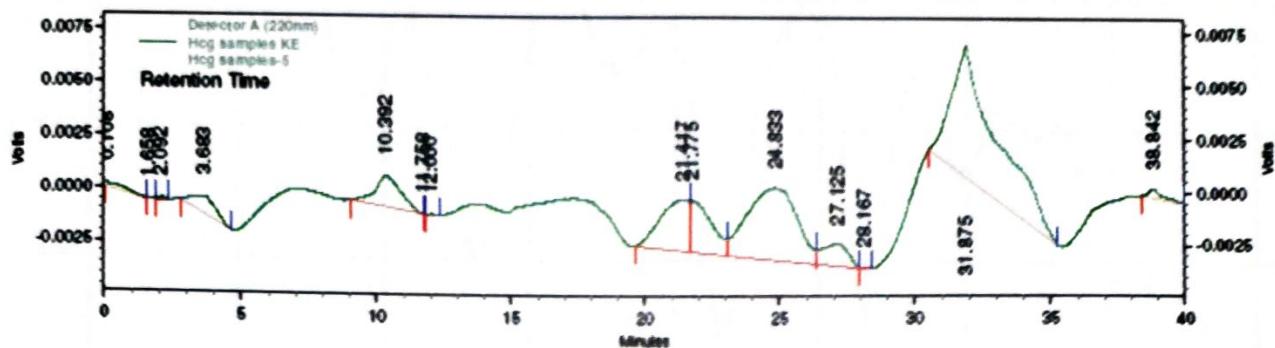
C:\Documents and Settings\gcms\Bureaublad\UHMC\Hcg samples-5, Detector A (220nm)

## Peak Area Calculation Sample KE

Shimadzu CLASS-VP V6.12 SP4  
Page 1 of 1

*Area % Report*

Method Name: C:\HPLC ANALYSIS\HCG\DATA\Hcg analysis.met  
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### Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.108	10778	0.638	238	1.276
2	1.658	380	0.022	44	0.236
3	2.092	1347	0.080	103	0.552
4	3.683	46830	2.770	836	4.483
5	10.392	94390	5.583	1447	7.760
6	11.758	155	0.009	41	0.220
7	12.000	397	0.023	31	0.166
8	21.417	176487	10.439	2410	12.924
9	21.775	131857	7.799	2404	12.892
10	24.833	433107	25.618	3393	18.196
11	27.125	68193	4.034	1057	5.668
12	28.167	599	0.035	43	0.231
13	31.875	712097	42.120	6181	33.147
14	38.842	14035	0.830	419	2.247

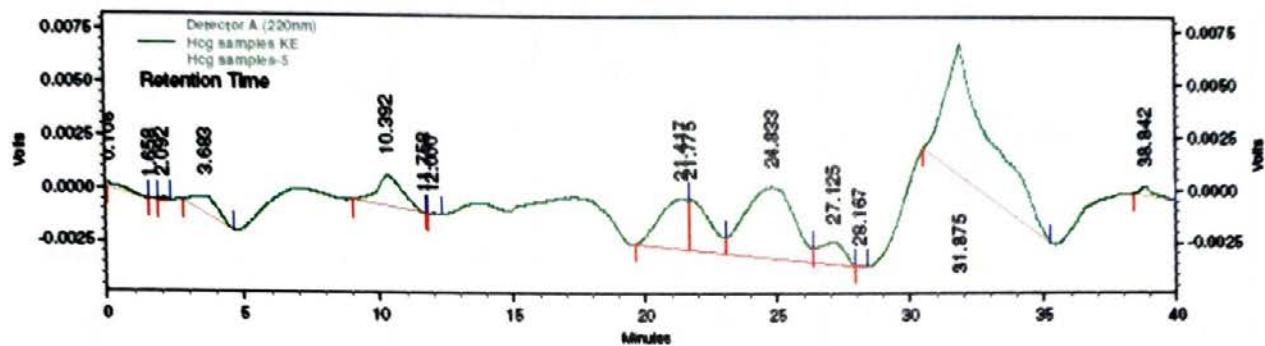
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## Peak Area Calculation Sample KE

Shimadzu CLASS-VP V6.12 SP4  
Page 1 of 1

*Area % Report*

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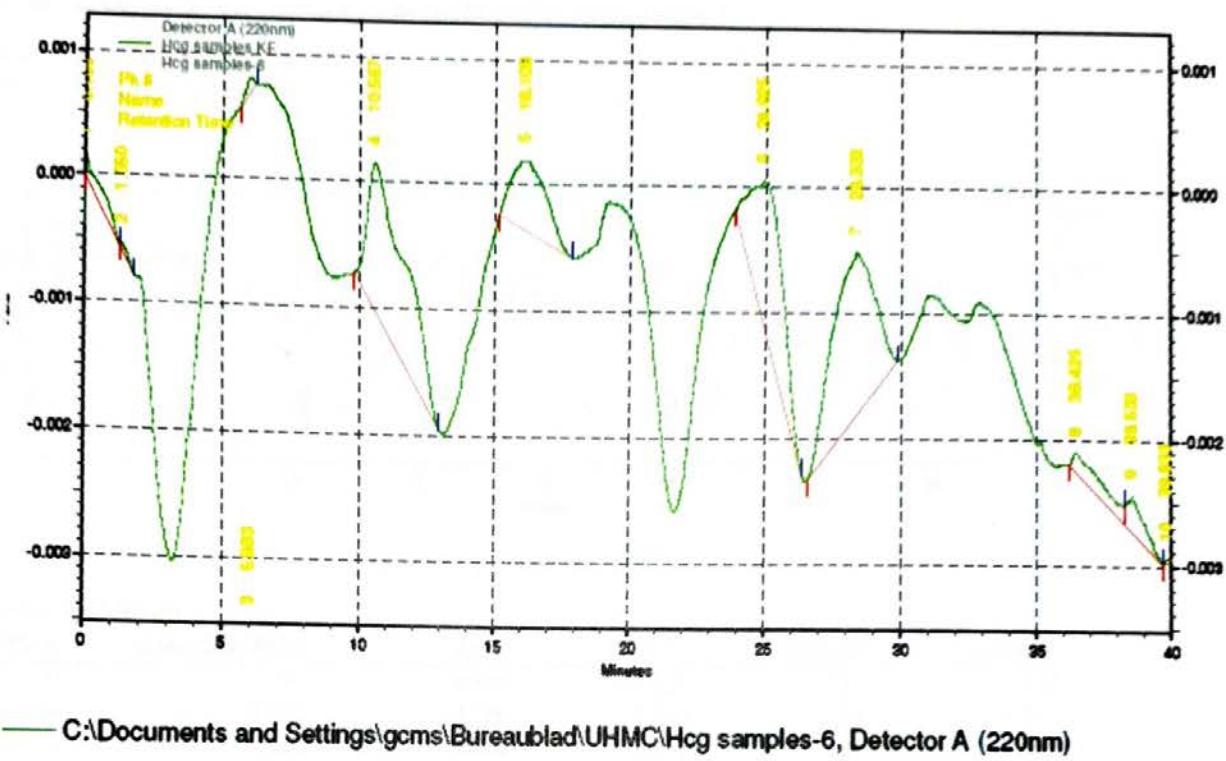
Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.108	10778	0.638	238	1.276
2	1.658	380	0.022	44	0.236
3	2.092	1347	0.080	103	0.552
4	3.683	46830	2.770	836	4.483
5	10.392	94390	5.583	1447	7.760
6	11.758	155	0.009	41	0.220
7	12.000	397	0.023	31	0.166
8	21.417	176487	10.439	2410	12.924
9	21.775	131857	7.799	2404	12.892
10	24.833	433107	25.618	3393	18.196
11	27.125	68193	4.034	1057	5.668
12	28.167	599	0.035	43	0.231
13	31.875	712097	42.120	6181	33.147
14	38.842	14035	0.830	419	2.247

Totals		1690652	100.000	18647	100.000
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#### 4.6 Results Sample KF

HPLC Chromatogram Sample KF

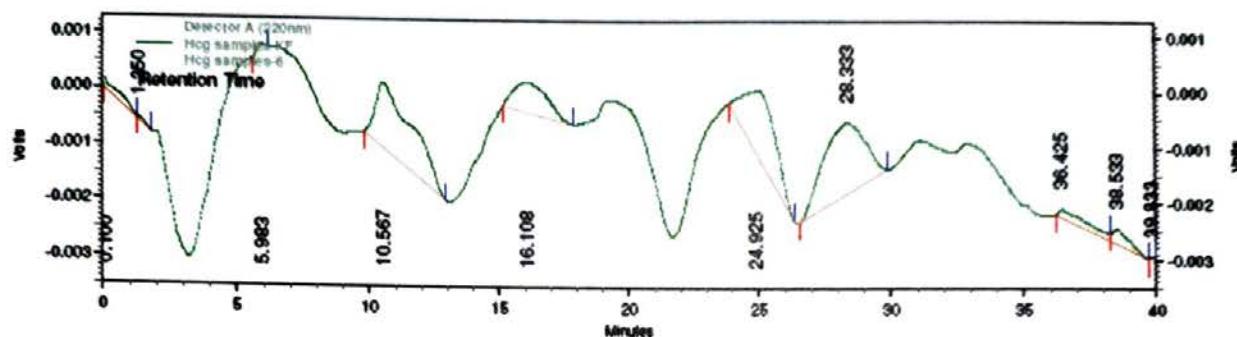


## Peak Area Calculation Sample KF

Shimadzu CLASS-VP V6.12 SP4  
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*Area % Report*

Method Name: CAHPLC ANALYSIS\HCG\DATA\Hcg analysis.met  
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### Detector A (220nm)

Pk #	Retention Time	Area	Area %	Height	Height %
1	0.100	8348	1.838	191	3.825
2	1.350	1196	0.263	50	1.001
3	5.983	2420	0.533	129	2.583
4	10.567	120804	26.597	1161	23.248
5	16.108	49541	10.907	549	10.993
6	24.925	101718	22.395	1152	23.068
7	28.333	142448	31.362	1296	25.951
8	36.425	15143	3.334	151	3.024
9	38.533	12242	2.695	282	5.647
10	39.833	339	0.075	33	0.661

Totals		454199	100.000	4994	100.000
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## **5.0 INTERPRETATION OF LABORATORY ANALYTICAL RESULTS**

Following the Laboratory assessment, the following samples were found to contain human chorionic gonadotropin (HCG):

- Sample KA retention time (RT) 36.283 and peak area of 37.539%
- Sample KB retention time (RT) 35.825 and peak area 26.512%
- Sample KC retention time (RT) 38.042, peak area 23.939

All the other samples analysed did not exhibit significant elution after 35 mins and the samples could be free from HCG.